Abstracts for the 19th Annual Meeting of the American College of Toxicology

November 8-11, 1998

SYMPOSIUM 1

Human Corneal Epithelial Cell Lines as in vitro Models for Evaluating Eye Irritation

F. H. Kruszewski, T. L. Walker, S. L. Ward, and L. C. DiPasquale, The Gillette Company, Gillette Medical Evaluation Laboratories, Gaithersburg, MD

Stratified cultures of transfected human corneal epithelial cell lines, HCE-T cells, have been grown on permeable membranes to create the "HCE-T Model." This system is a tissue equivalent of the corneal surface that will support the ocular toxicology testing of consumer products. The Model is maintained in a format where it is basally exposed to media and apically exposed to air. In this format, cells stratify into four to six cell layers, express tight junctions, produce an apical layer of mucin, and demonstrate a barrier property, all which are features similar to the corneal epithelium. The barrier function of the HCE-T Model can be demonstrated in vitro by its ability to retain sodium fluorescein and to maintain a significant electrical resistance. Transepithelial permeability to sodium fluorescein (TEP) has been used in this model as a physiologically relevant, standard assay to screen for the ocular irritancy potential of water-soluble substances. This protocol involves the generation of concentration-response data for test materials. Groups of 20 diverse chemicals and 25 surfactant-based product formulations have been tested in the HCE-T TEP assay, and in vitro data was compared to historical Draize rabbit eye irritation test maximum average scores (MAS). The Pearson's correlation coefficient, r. for these comparisons were 0.71 and 0.86 respectively. A separate protocol for evaluating the ocular irritancy potential of water-insoluble substances in the HCE-T Model is in development. The TEP assay is not applicable since these samples generally can't be diluted in aqueous diluents nor reliably washed from the culture surface. This alternate protocol involves the generation of time-response data for test materials. Toxicity endpoints that have been evaluated through assay of basal media were changes in the inflammatory mediator, prostaglandin E₂, and the metabolite, lactate. Preliminary data from the analysis of four formulations indicated that only the lactate assay results corresponded sufficiently to the historical Draize MAS to suggest the possible utility of this endpoint. Further protocol research and database development will focus on providing a battery of useful assays to support the ocular toxicological analysis of a wide array of chemicals and with varied physical forms.

Strategic Use of the Stress Gene Heme Oxygenase-1 as an Index in Evaluating and Screening Drugs and Chemicals for Ocular Formulations

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Purpose. Predicting the toxic potential of compounds to the ocular surface has been dependent on the Draize test for the past half century. Alternatives to Draize testing have recently been sought. Stress gene expression has emerged as a means of quantifying cellular reaction and, thus, the toxic potential of the compound in question. This study examines the expression of the major stress response gene heme oxygenase-1 (HO-1) in a human transfected corneal epithelial cell (HCE-T) line following challenges with a number of known ocular irritants and evaluates the effectiveness of using the HO-1 promoter activation as a stress index.

Methods. Irritants tested included hydrogen peroxide, isopropyl alcohol, sodium hydroxide, Tween 20 trichloroacetic acid, triclosan, aluminum chloride and SDS. HCE-T cells were grown to 80% confluency and treated with the listed irritants at a concentration ranges of 10–100 μM. Cell viability, Northerm blot analysis and RT/PCR were performed following a 1–48 hrsincubation period. HCE-T cells were transfected with the HO-1 promoter/CAT or Luciferase constructs and CAT/Luciferase activity was measured 1–12 hrs after irritant treatment.

Results. Northern blot analysis demonstrated that levels of HO-1 mRNA transcripts increased regularly after exposure to the irritants in a concentration dependent manner. Quantitative RT/PCR confirmed the validity of the Northern analysis results and showed direct relationship to cell viability. Transient transfection of HO-1 promoter CAT/or Luciferase constructs into HCE-T demonstrated that these irritants transcriptionally activate HO-1 gene and the degree of activation correlated with the toxicity of the irritant.

Conclusions. These results are consistent with the idea that HO-1 mRNA levels may be used as an indicator of irritant toxicity and that HCE-T cells respond to stress in a fashion similar to other human cell lines. This strategy for testing will enable us to

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